

Repetitive Reaction and Restitution (R³) Induction of Drought Hardiness in Conifer Container Seedlings

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Abstract: Planting failures are often attributed to unexpectedly harsh conditions after planting. Characterization of soil water at the planting site, along with associated influences of site preparation and soil texture, is recommended. Additionally, tree planting technique and seedling biology should be targeted to site conditions.

A nursery regime for induction of drought hardening is proposed for hot-lift summer- and fall-planted container conifers.

Keywords: seedling hardiness, nursery culturing, drought stress

Introduction

Nursery growers, researchers, and field foresters have improved forest seedling quality, planting technology, and site preparation with a corresponding increase in survival and growth after outplanting. This expertise has been disseminated worldwide through publications and conferences. In the 1970s and 1980s, average survival for planted seedlings was 50-75% (Cayford 1978; Mexal and others 2008). Today we expect survival of 90% or greater, although particularly harsh sites may need to be planted more than once due to poor seedling survival where water is a limiting factor (Mexal and others 2008; Regan and Davis 2008).

Characterizing the Site

Site preparation treatments such as vegetation control, scarification, and cultivation, along with annual weather patterns, result in a range of site conditions for outplanting. Lantz (1996) describes North Carolina standards for planting categories as normal, critical, and severe based temperature (Table 1), relative humidity, wind, and soil moisture on the day of planting. To further characterize the site, soil texture must be considered; the ability to hold water varies among soils (Table 2; Figure 1). When soil water potential is below -60 to -70kPa (-0.6 to -0.7 bar), root elongation ceases (Grossnickle 2005). In order to characterize the planting site for outplanting dates beyond the traditional outplanting windows, a soil moisture stress curve should be estimated based on a progression of moisture conditions at selected depths (Figure 2) using standard methods for determining soil hydraulic conductivity. This curve will suggest the moisture stress level likely to be encountered by a newly planted seedling before new root growth penetrates to a new source of moisture.

Table 1. North Carolina planting categories (Lantz 1996).

	Normal	Critical	Severe
Temperature	2 to 24 °C (35 to 75 °F)	24 to 29 °C (76 to 85 °F)	> 29 °C (> 85 °F)
Relative Humidity	>50%	30-50%	<30%
Wind	<10mph	>10mph	>15mph
Soil moisture buildup	<30	30-80	>80

Table 2. Storage capacity for different soil texture classes (inches water/feet depth). (1 in = 2.54 cm; 1 ft = 0.3 m)

Texture class	Field capacity	Permanent wilting point
Sand	1.2	0.3
Fine sand	1.4	0.4
Sandy loams	1.9	0.6
Fine sandy loams	2.6	0.8
Loams	3.2	1.2
Silt loams	3.4	1.4
Light clay loam	3.6	1.6
Clay loam	3.8	1.8
Heavy clay loam	3.9	2.1
Clay	3.9	2.5

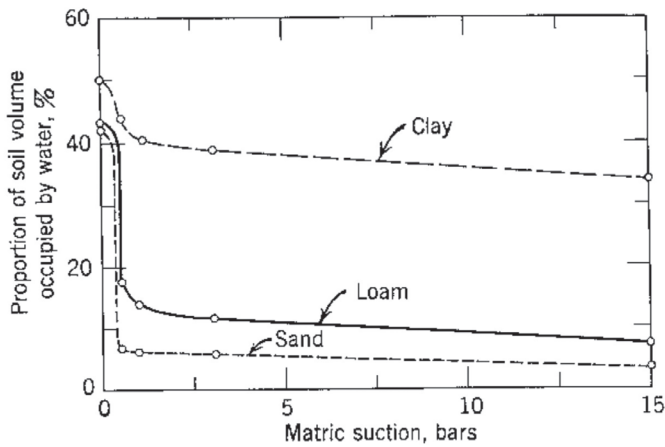


Figure 1. Volumetric water content of three soils at different matric suctions (from Black 1968).

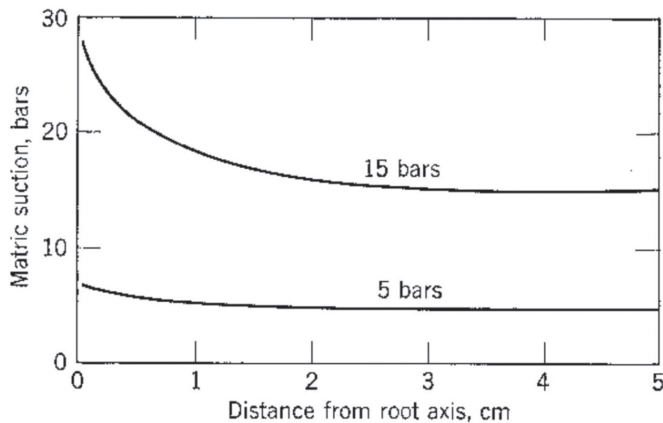


Figure 2. Matric suction versus distance from root in bulk sandy loam soil at -5 and -15 bars (Black 1968).

The Root/Soil Interface

Kramer and Kozlowski (1960) note that pines (*Pinus* spp.) have few growing root tips during winter, yet can absorb considerable quantities of water. Suberized roots can be a major route of water uptake in tree root systems, through lenticels on the surface of the root and by discontinuities in the periderm (bark) plates although unsuberized (white) root tips on growing roots have the highest uptake per unit of surface area (Carlson and others 1990). New root growth leads to an increase in daily minimum water potential (Ψ) for newly planted seedlings (Figure 3) (Grossnickle 2005). Given that the newly planted seedling may encounter a soil moisture stress condition that precludes new root growth, survival depends upon creating the water continuum with the existing root system—the hydrostatic system (Kramer and Koslowski 1960) or the soil-plant-atmosphere continuum (SPAC) (Grossnickle 2005).

Grossnickle (2005) provides an excellent discussion of the importance of root-soil contact in the planting of seedlings and points out that poor contact will increase the resistance to water movement from the soil to the plant. Landis and Dumroese (2009) emphasize the importance of ensuring the planting stock is fully hydrated at the time of planting. When the soil is dry or the soil texture is coarse, the South African “Puddle-Plant” technique may be employed, with the addition of 3 to 5 L (0.8 to 1.3 gal) of water per plant (Viero 2000). Seedling water demand in the first few days after outplanting can be estimated from irrigation studies in the nursery immediately before lifting. When outplanting takes place outside the traditional outplanting window, however, hydration at the time of outplanting may be insufficient to meet moisture needs until the next rainfall event. Water deficits can inhibit physiological processes such as photosynthesis with a consequent reduction in root growth (Joly 1985).

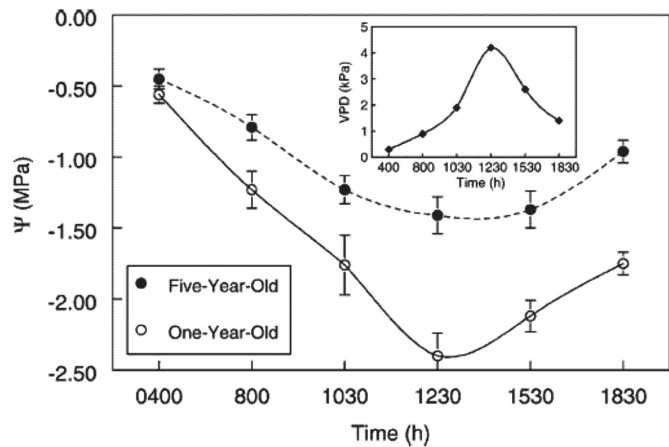


Figure 3. Diurnal Ψ of newly planted and established lodgepole pine seedlings. Insert of vapor pressure deficit (Grossnickle 2005).

Water uptake can occur via an apoplastic or a symplastic pathway (Figure 4), both of which are influenced by solute concentrations and osmosis. As the distance from the root increases, soil moisture can be expected to increase. Water content in close proximity to the roots, however, can be increased due to production of mucigel. The peripheral cells of the root cap and the epidermal cells of the root produce and secrete large amounts of mucigel, a slimy substance made by dictyosomes. Mucigel is a hydrated polysaccharide containing sugars, organic acids, vitamins, enzymes, and amino acids (Walker and others 2003; Earley 2010). A prominent role of mucigel is to provide continuity between root surfaces and soil moisture.

Proposed R³ Protocol _____

In the nursery, mild moisture stress (-1.5 MPa [-15 bars]) is often used to induce dormancy, but foliar injury can occur at higher levels (-1.8 to -2.0 MPa [-18 to -20 bars]) (Landis 1999). Lopushinsky (1990) notes that drought hardiness can be increased by exposure to moderate water stresses (-0.5 to -1.0 MPa [-5 to -10 bars]) and cites work by Christersson showing that pot-grown Scots pine (*Pinus silvestris* L.) and Norway spruce (*Picea abies* [L] Karst.) seedlings could increase drought tolerance to -3.5 MPa (-35 bars), compared to -2.5 MPa (-25 bars) for unhardened seedlings. Joly (1985) suggested that the turgor pressure that leads to observable wilt probably changes as a result of water stress conditioning, and Stocker (1960) describes the “time course of changes in the plasmatic viscosity” as comprising two phases—Reaction phase (destruction of the plasma) and Restitution phase (reparation of the damage).

Reaction Phase

At the beginning of the hardening regime, seedlings are leached with water (to reduce growing medium solution EC) and then dried down to the “Just Open Limit” (50% stomatal closure; temporary wilting). This increases respiration, decreases transpiration and photosynthesis, and reduces growth. The expected ψ is approximately -2500 kPa on these temporarily wilted seedlings.

Restitution Phase

Seedlings are re-wetted with hardener fertigation. The irrigation system must be adequate to cool the entire crop within minutes. This increases photosynthetic rate. This regime is continued for 1 to 2 weeks.

Repetition²

Reaction and Restitution phases are repeated. The temporary wilt will likely be observed on those seedlings that did not wilt on the first dry-down. Photosynthetic capacity may become greater after successive wilting periods (Stocker 1960).

Repetition³

Reaction and Restitution phases are repeated again. The temporary wilt will likely not be observed on this final dry down.

Once the steps have been completed, seedlings are re-wetted with nutrient loading solution, then harvested and shipped.

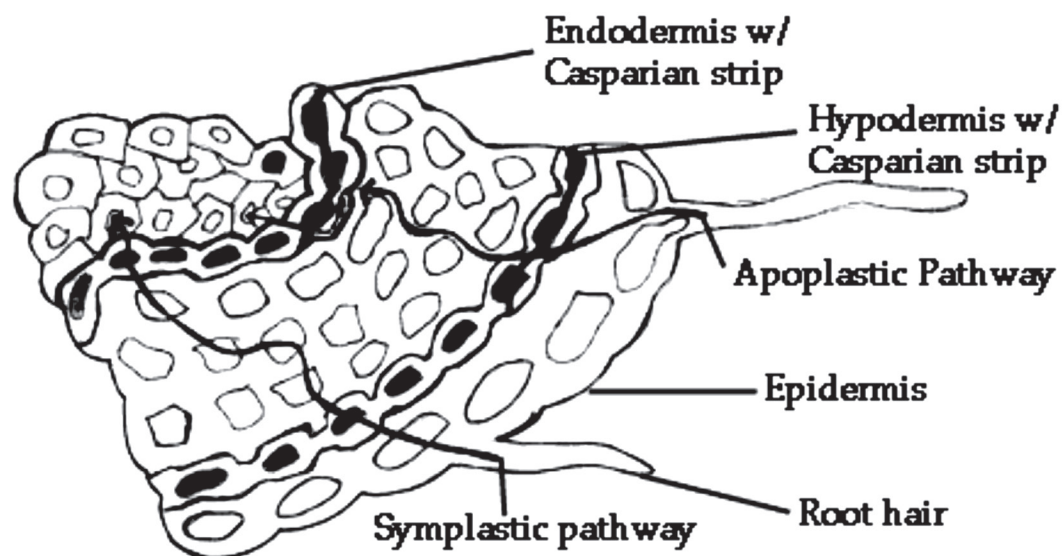


Figure 4. Water uptake pathways (Earley 2010).

Similar drought pre-conditioning protocols have been investigated over the past several decades with varying results. Any change to culturing regimes should first be tried on a small scale for each species, seed zone, and stocktype in question to determine its influence on subsequent crop quality and outplanting performance.

References

- Black CA. 1968. Soil-plant relationships. New York (NY): John Wiley and Sons. 792 p.
- Carlson WC, Miller DE. 1990. Target seedling root system size, hydraulic conductivity, and water use during seedling establishment. In: Rose R, Campbell SJ, Landis TD, editors. 1990. Target seedling symposium: proceedings, combined meeting of the western forest nursery associations; 13-17 August 1990; Roseburg, OR. Fort Collins (CO): USDA Forest Service, Rocky Mountain Forest and Range Experiment Station. General Technical Report RM-200. p 53-65.
- Cayford JH. 1978. Forest regeneration in Canada—at the crossroads. In: van Eerden E, Kinghorn JM, editors. Proceedings of the root form of planted trees symposium; 16-19 May 1978; Victoria, BC. Victoria (BC): Pacific Forest Research Centre, Canadian Forest Service, Fisheries and Environment Canada. Joint Report Number 8.
- Earley E. 2010. Roots. URL: www.cartage.org.lb/en/themes/sciences/botanicalsciences/plantsstructure/roots/roots.htm (accessed 2 July 2010).
- Grossnickle SC. 2005. Importance of root growth in overcoming planting stress. *New Forests* 30:273-294.
- Joly RJ. 1985. Techniques for determining seedling water status and their effectiveness in assessing stress. In: Duryea ML, editor. Evaluating seedling quality: principles, procedures, and predictive abilities of major tests; 16-18 October 1984; Corvallis, OR. Corvallis (OR): Oregon State University, Forest Research Laboratory.
- Kramer PJ, Kozlowski TT. 1960. Physiology of trees. New York (NY): McGraw-Hill. 642 p.
- Landis TD. 1999. Seedling development: the establishment, rapid growth, and hardening phases. In: Landis TD, Tinus RW, McDonald SE, Barnett JP. The container tree nursery manual. Volume 4. Washington (DC): USDA Forest Service. Agricultural Handbook 674.
- Landis TD, Dumroese RK. 2009. A visit to Finland—an opportunity to see with “new eyes.” *Forest Nursery Notes* 29(1).
- Lantz CW. 1996. A guide to the care and planting of southern pine seedlings. Atlanta (GA): USDA Forest Service Southern Research Station. Management Bulletin R8 – MB39.
- Lopushinsky W. 1990. Seedling moisture status. In: Rose R, Campbell SJ, Landis TD, editors. 1990. Target seedling symposium: proceedings, combined meeting of the western forest nursery associations; 13-17 August 1990; Roseburg, OR. Fort Collins (CO): USDA Forest Service, Rocky Mountain Forest and Range Experiment Station. General Technical Report RM-200. p 123-138.
- Mexal JG, Rangel RAC, Landis TD. 2008. Reforestation success in central Mexico: factors determining survival and early growth. *Tree Planters Notes* 53(1).
- Regan DJ, Davis AS. 2008. Effect of container size and copper treatment on western white pine seedling growth and survival. In: Dumroese RK, Riley LE, technical coordinators. National Proceedings: Forest and Conservation Nursery Associations—2007. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. Proceedings RMRS-P-57. P 104-108.
- Stocker O. 1960. Physiological and morphological changes in plants due to water deficiency. *Plant-Water Relationships in Arid and Semi-Arid Conditions*, UNESCO, Arid Zone Restoration 15:63-104.
- Viero P. 1990. Planting. In: Owen DL, editor. *South Africa Forestry Handbook 2000*. Volume 1. Pretoria (South Africa): The Southern Africa Institute of Forestry.
- Walker TZ, HP Bais, E Grotewold, JM Vivanco. 2003. Root exudation and rhizosphere biology. *Plant Physiology* 132:44-51.